

Chapter 3 Exploring Proteins and Proteomes

The Proteome is the Functional Representation of the Genome p. 66

The proteome the combination of the functional proteins in a cell.
The genes that have been expression

3.1 The Purification of Proteins is an Essential First Step in Understanding Their Function pp. 66-78

"Never waste pure thoughts on an impure protein"

The Assay: How do We recognize the Protein That We Are Looking For? p. 67

When you want to purify a protein you have to know when you have it.

The way we recognize this is called an assay

With enzymes you can assay their activity

Usually given in enzyme units (eu)

generally $\frac{\mu\text{mol}}{\text{min}}$ or $\frac{\text{mmol}}{\text{min}}$

Sometimes you can directly measure the reaction

$\text{NADH} \rightarrow \text{NAD}^+$
- lactate dehydrogenase

Radioactive assay

Protein Assays - detect protein concentration

BioRad assay

Lowry assay

280 nm

Units in mg

Specific Activity

activity / protein

Unit - eu/mg

rises as purification proceeds

Object of purification is to maximize the specific activity

Proteins Must Be Released From the Cell to be Purified pp. 67-68

Break the cell open

*French press
Chemical lysis
sonication*

Get homogenate

Centrifugation

Spin down after breaking cells

IF your protein is soluble just on high speed spin

IF protein is the insoluble or in a membrane

Differential centrifugation

Spin harder & harder, Collect pellets which the protein is in

Proteins Can Be Purified According to Solubility, Size, Charge, and Binding Affinity pp. 68-71

Purification according to solubility size, charge, and binding affinity

Salting out

Proteins are in general less soluble at a high salt conc.
Ammonium sulfate

Dialysis

Slide Proteins can be separated from small molecules through dialysis

Esp. good for getting rid of salt

Small molecules will pass out through the pores of the dialysis tubing

Gel-filtration chromatography

Slide separates proteins based on size

beads with tunnels

larger proteins come out first

playground example

Ion exchange chromatography

Slide separates protein on charge

pI's are important

negative proteins stick to positive columns - anion exchange

positive proteins stick to negative columns - cation exchange

Increasing salt washes out protein

Hydrophobic chromatography (not in book)

Slide Binds to non-polar patches

start at high [salt] and lower

Affinity chromatography - Best if you can find one that will work

~~Best~~ Takes ~~advantage~~ advantage of affinity some proteins have for certain chemicals

Maybe attach a molecule the protein binds to the column

conoclain A + glucose

Nickel columns

High-Pressure Liquid Chromatography

- uses other types of columns

- uses finer beads so needs high pressure to run

- finer beads leads to better separation

Proteins Can be Separated by Gel Electrophoresis and Displayed pp. 71-74

Not purification - Evaluation of purification!

How can we tell if our purification worked?

1. Gel electrophoresis

2. Increasing Specific activity

Gel electrophoresis

Slide Separates macromolecules based on size & charge

Separates them in an electric field

Homework Question

Common gels are polyacrylamide & agarose

Slide The are large polymers... like one big size exclusion beads
Have holes in them that smaller molecules can get through
Big molecules left behind

Question Proteins usually separated by SDS-PAGE

Slide SDS - detergent that denatures the protein & coats them in (-) charge

BME or dithiothreitol breaks disulfide bonds

- roughly 1 SDS molecule per a.a. so the charge is ok to the size of the protein

Slide Separates the proteins on size only

To visualize coomassie blue, silver stain, or radiolabeled

Coomassie blue detects to 0.1 μ g

silver stain can detect to 0.02 μ g

Slide Isoelectric focusing

separates proteins based on pI

passes proteins through a gel w/ a pH gradient

polyampholytes are passed through the gel

The protein stops at its pI

Slide Two dimensional electrophoresis

Combines SDS-PAGE w/ isoelectric focusing

First IF

Then SDS-PAGE

Slide Good for when looking for in a large combination

A Protein Purification Scheme Can Be Quantitatively Evaluated pp. 74-75

Protein purification evaluation

Total Protein

Slide Total activity

Specific Activity

Slide Yield (total remaining activity)

Purification level (times SA has increased)

Need to have good yield and specific activity

Ultracentrifugation is Valuable for Separating Biomolecules and Determining Their Masses pp. 76-77

Slide

When being centrifuged molecules will move through the medium depending on several factors.

Slide

1. The mass – more mass moves through quicker
2. Shape – because of drag – elongated moves slower than spherical
3. Density of the molecule – more dense move faster (combo of 1 & 2)
4. Density of a solution – if the molecule is less dense than the sol'n they won't move through

Sedimentation coefficients are unique to molecules. Usually is Svedberg constant (S)
- seen w/ Ribosomes (5S, 16S, etc.)

3.2 Amino Acid Sequences Can Be Determined by Automated Edman Degradation pp. 78-84

Proteins can be sequenced using Edman Degradation Edman Degradation

Protein sequencing is not as important b/c the advent of recombinant DNA technology

DNA is easier to sequence + we get protein from DNA
Doesn't show post translational modification

3.3 Immunology Provides Important Techniques with Which to Investigate Proteins pp. 84-90

Antibodies to Specific Proteins Can be Generated

Slide Antibody (Immunoglobulin) – protein that binds invaders of the organism

Slide Antigen – invader (protein, nucleic acids, polysaccharides)

The binding of the Antibody to the antigen is the beginning of the immune response

Antigenic Determinant or Epitope – specific area recognized on antigen

Making Antibodies